

# Evaluation of a Multiplexed, Sample-to-Answer, Microarray System for the Detection of Encephalitic Viruses

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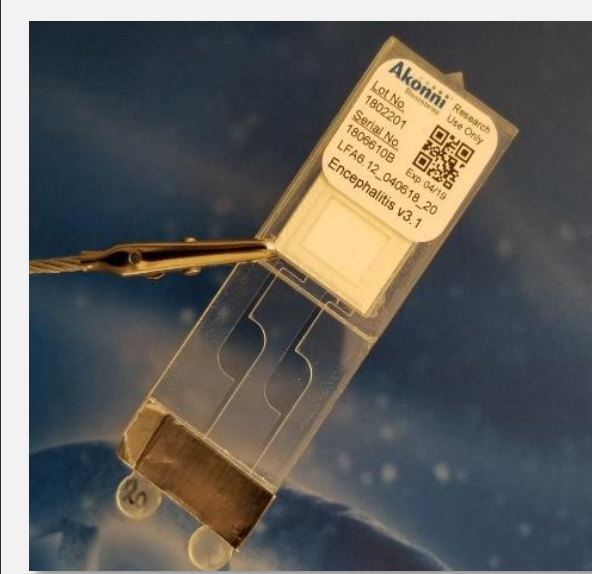
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## Background

- Identification of the cause of encephalitis can prove challenging as cerebrospinal fluid (CSF) is an invasive specimen to collect, with small sample volumes and commonly low viral titers.
- Molecular assays, with high sensitivity and the potential for multiplexing are advantageous.
- We developed an encephalitis microarray platform in an all-plastic lateral flow cell, for the detection of HSV1, HSV2, VZV, CMV, HHV6, enterovirus, and West Nile virus (WNV).
- Lateral flow cell designed for simplified workflow, including simultaneous amplification and microarray detection within a single reaction chamber, and in a fully contained, closed amplicon system.
- The analytical and clinical sensitivity of this assay was evaluated from extraction, through amplification, hybridization, and imaging, to assess its performance relative to that of individual real-time assays for the same targets.

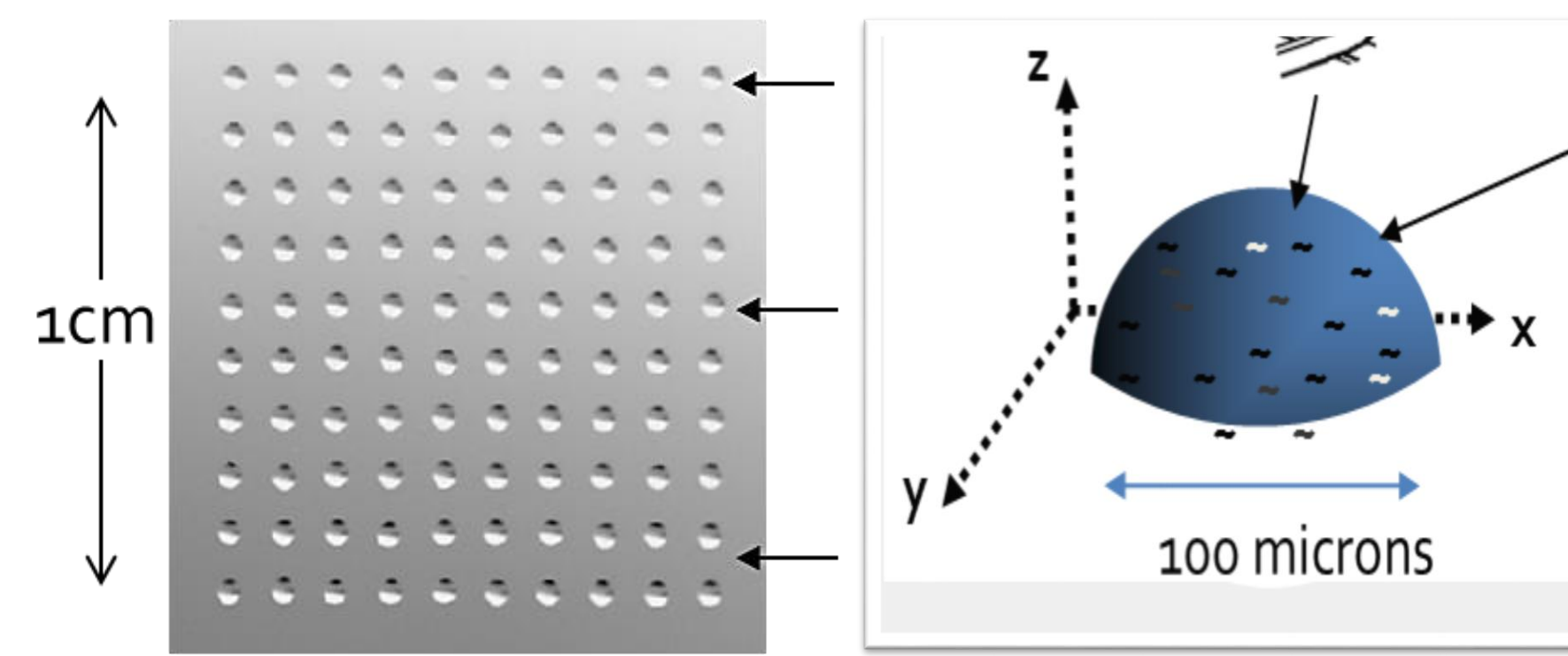
## Akonni Biosystems Encephalitis TruArray Panel



- Lateral Flow Array (LFA) – Plastic slide containing two flow chambers houses low-density “gel-drop” microarrays
- Gel elements are copolymerized with specific target capture probe and have a 3D structure allowing increased hybridization efficiency
- Amplification, hybridization, and imaging all occur onboard the LFA within a closed system

## 8-Plex Primer Pool

Target	Gene Target
HSV1	UL44 Envelope glycoprotein C
HSV2	UL3 Nucleoprotein
VZV	ORF28: DNA Polymerase catalytic subunit
CMV	UL54 DNA Polymerase
HHV6	U38 DNA Polymerase
Enterovirus	5' UTR
WNV	Envelope
GFP	GFP Transcript



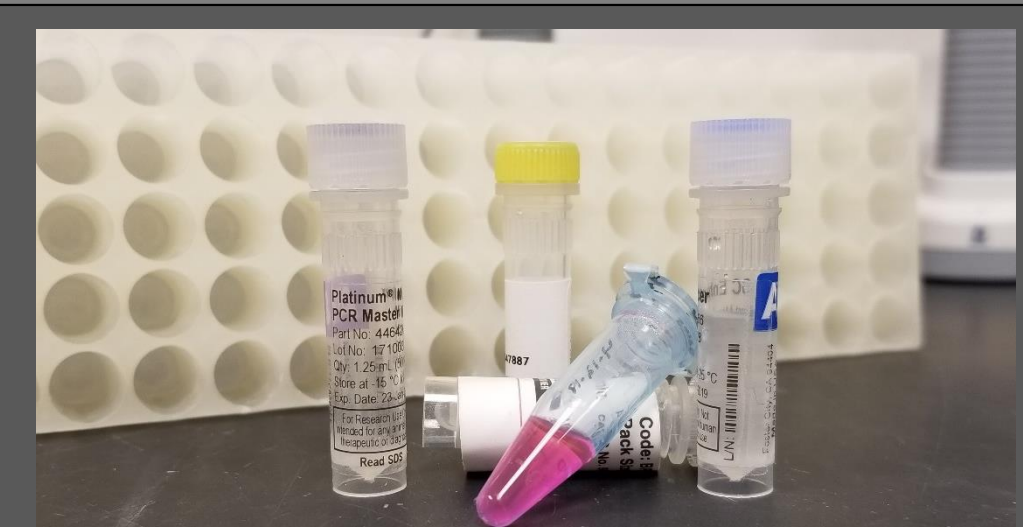
## Methods

### Extraction



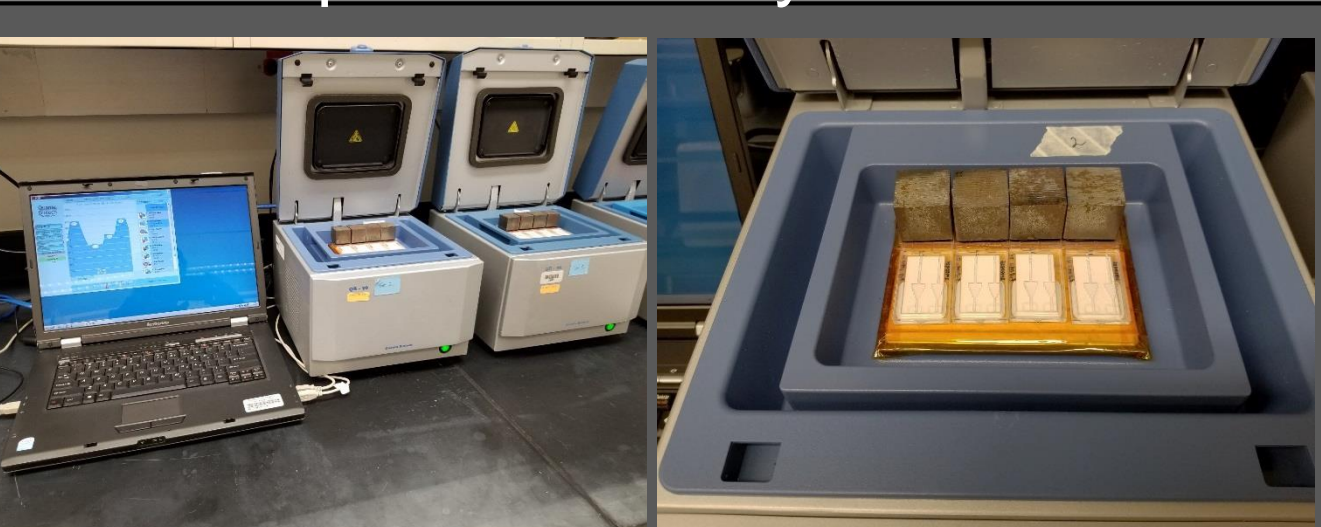
- Artificial CSF amended with quantified viral isolates
- Serially diluted, divided into multiple replicates
- Spiked with GFP RNA transcript as an extraction, amplification, and inhibition control.
- Extraction and onboard reverse transcription performed using Akonni TruTip Technology on Automated Work Station

### Master mix



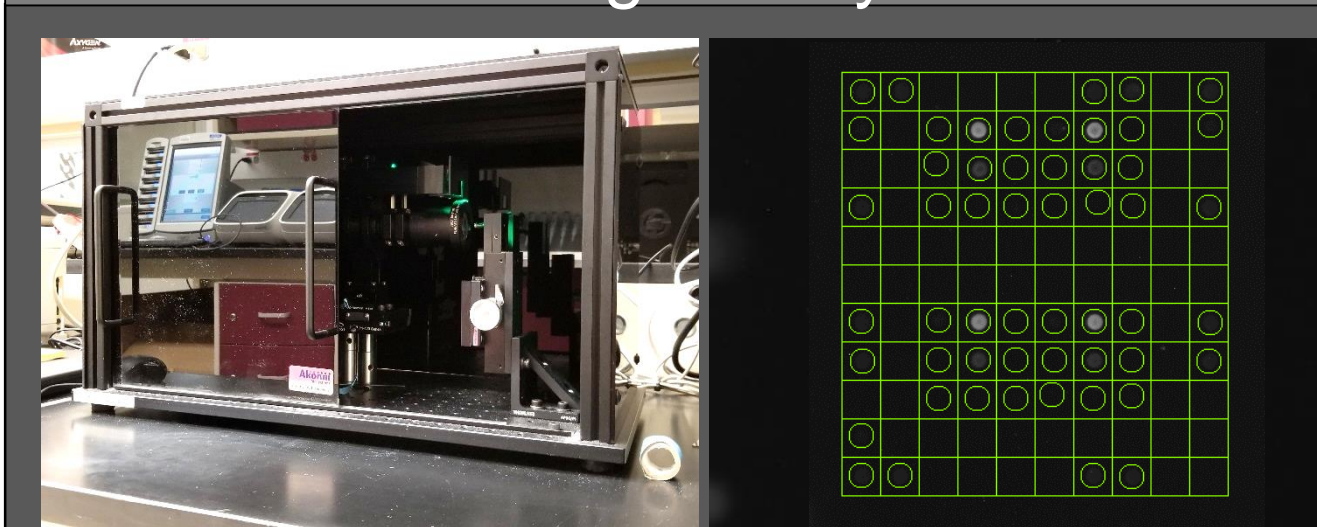
- Amplification and hybridization with Platinum Multiplex PCR Master mix (ThermoFisher).
- Master mix for encephalitis TruArray Panel utilizes an 8-plex primer pool containing self-avoiding molecular recognition system (SAMRS) primers for all targets except VZV

### Amplification & Hybridization



- Quantified RT products amplified and hybridized simultaneously on plastic lateral flow cell microarrays
- Asymmetric PCR with 10:1 primer ratio. 10X primers are Cy-3 labeled for single-stranded amplicon binding and imaging
- Performed on heat blocks, modified with a copper strip to isolate heat transfer to LFA.

### Image Analysis



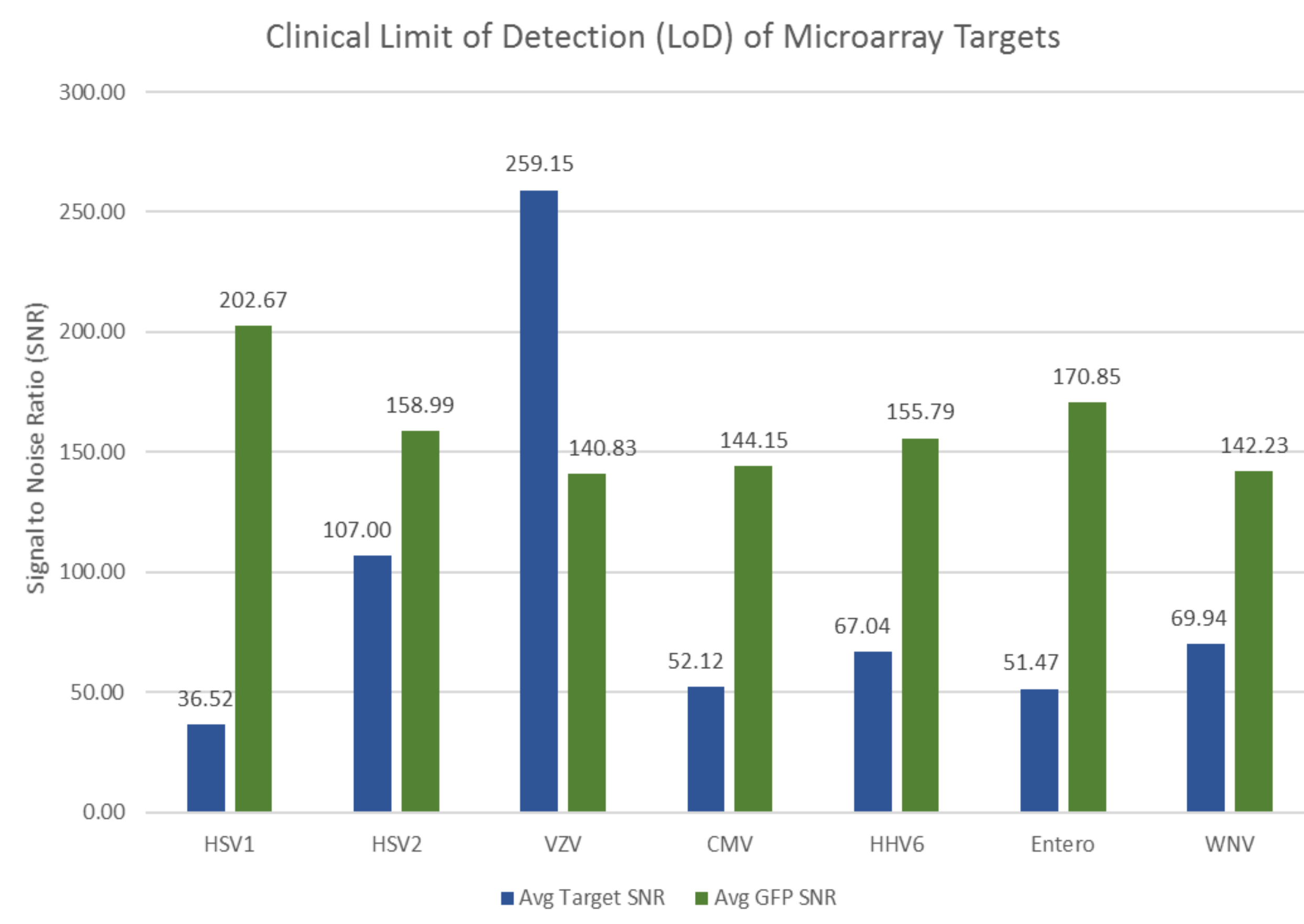
- LFA's washed and imaged directly on the slide with 1000ms exposure
- Amplicon signal is measured against background to generate a signal to noise ratio (SNR) for each gel spot

## Encephalitis TruArray v.3.1 Chip Map



- (Left) Sample loading and LFA washing conducted through a single port onboard the array
- (Right) Example of HHV6 Positive microarray
  - Targets are measured in quadruplicate per microarray
  - Green Fluorescent Protein (GFP) used as internal positive control
  - Cy3 beacons on microarray used for positional reference

## Clinical Sensitivity



Viral Target	HSV1	HSV2	VZV	CMV	HHV6	Enterovirus	WNV
<b>Avg Target SNR</b>	36.52	107.00	259.15	52.12	67.04	51.47	69.94
<b>Avg GFP SNR</b>	202.67	158.99	140.83	144.15	155.79	170.85	142.23
<b>Dilution gc/ml</b>	1.03E+04	5.21E+04	1.62E+03	6.55E+03	1.28E+04	1.05E+04	1.07E+04
<b>Average gc/ul</b>	14	56	8	10	9	8	3
<b>Average gc/rxn</b>	58	242	36	43	40	36	13
<b>Average CT</b>	35.18	33.42	37.11	33.15	31.43	33.35	33.45
<b># Positive</b>	39/40	40/40	40/40	40/40	40/40	40/40	55/56
<b>% Positive</b>	97.50%	100%	100%	100%	100%	100%	98.21%

- Limits of detection (LoD) – determined as lowest viral DNA concentration in which at least 95% of extracted replicates were detected on the array (starting from extraction)
- Quantitation of virus determined by real-time PCR standard curves using in-house developed standards

## Analytical Sensitivity

Viral Target	HSV1	HSV2	VZV	CMV	HHV6	Enterovirus	WNV
<b>Avg SNR</b>	45	43	300	178	25	65	63
<b>GC/rxn</b>	90.00	195	< 25	43	22	4	5
<b>SNR range</b>	42-48	3-107	262-327	114-252	3-47	44-86	31-95
<b># Positive</b>	2/2	8/8	4/4	4/4	4/4	2/2	2/2

- Analytical sensitivity – determined as lowest viral DNA concentration in which all replicates tested positive
- Determined using nucleic acid from high titer in-house controls

## Analytical & Clinical LoD Comparison

Viral Target	Analytical Sensitivity (gc/rxn)	Clinical Sensitivity (gc/rxn)
HSV1	90	58
HSV2	195	242
VZV	< 25	36
CMV	43	43
HHV6	22	40
Enterovirus	4	36
WNV	5	16

- <sup>a</sup> Detection of reverse transcribed product from viral nucleic acid
- <sup>b</sup> Detection of reverse transcribed product after automated TruTip extraction and RT of virus-amended samples

## Conclusions/Discussion

- A multiplexed amplification microarray system has been developed for the detection of viral encephalitis targets in clinical CSF samples, with sensitivities below 50 gene copies/reaction for VZV, CMV, HHV6, enterovirus, and WNV in both analytical and clinical sensitivity experiments.
- Slightly higher detection limits of 60 and 200 gene copies/reaction were observed for HSV1 and HSV2 respectively.
- Additional optimization experiments are ongoing to improve the HSV1 and HSV2 sensitivity.
- The evaluation demonstrated the effective performance of all subparts of the fully integrated and automated sample-to-answer system, with high sensitivity and clinical utility in the multiplexed format.
- Further studies are planned to evaluate assay specificity, the effect of interfering substances, and array performance with clinical CSF samples.

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